

# **DOE Bioenergy Technologies Office (BETO) 2015 Project Peer Review**

## **Managing the Microbial Ecology of a Cyanobacteria- Based Photosynthetic Factory Direct!**

**Bruce E. Rittmann, Director  
Swette Center for Environmental Biotechnology  
Biodesign Institute at Arizona State University**

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Algae Technology Area Review

Principal Investigator  
Organization

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# Goal Statement

- Understand the factors controlling the productivity of a photobioreactor (PBR) that uses laurate-excreting *Synechocystis* in a Photosynthetic Factory Direct!
  - What controls laurate production by *Synechocystis*?
  - What controls laurate biodegradation by heterotrophic bacteria?
  - What strategies are needed to maximize net production?
- This work will lead to means to increase productivity and lower costs for microalgae-based biofuels.

# Quad Chart Overview

## Timeline

- December 1, 2012
- November 30, 2014
- 100% complete

## Barriers

- Barriers addressed
  - Low product yield
  - Poor crop protection
  - High product cost

## Budget

	Total Costs FY 10 – FY 12	FY 13 Costs	FY 14 Costs	Total Planned Funding (FY 15- Project End Date)
<b>DOE Funded</b>	00	218,037	250,597	27,965
<b>Project Cost Share (Comp.)*</b>	00	55,699	59,886	10,540

## Partners

- Partners
  - None

# 1 - Project Overview

- History
  - Originally a proposal submitted to NSF EFRI
  - Selected by DoE BETO for funding outside the NSF program
- Technical Direction
  - Enable the “Photosynthetic Factory Direct!”
  - Genetically modified *Synechocystis* sp. PCC6803 excretes laurate, a jet fuel precursor
  - We want to maximize the net laurate productivity by:
    - Understanding photosynthetic productivity by *Synechocystis*
    - Understanding heterotrophic bacteria that biodegrade laurate
    - Testing systems to maximize laurate capture and minimize its biodegradation

## 2 – Approach (Technical)

1. Fundamental studies on what wild-type and modified *Synechocystis* produce by photosynthesis
  - Organic-C partitioning – biomass, SMP, laurate
  - Kinetic control – light, nutrients, pH
2. Identification and characterization of prevalent strains of heterotrophic bacteria in *Synechocystis*-based PBRs
  - Phylogenetic diversity
  - Metabolic capability
  - Kinetics
3. Practical PBR strategy to minimize heterotrophs and improve laurate harvest

## 2 – Approach (Management)

- Study carried out totally within the PBR team of the Swette Center for Environmental Biotechnology
- PI – Bruce Rittmann
- Co-PI – Rosa Krajmalnik-Brown
- PhD Students – Alexander Zevin and Binh Nguyen
- Research Technologist – Megha Patel
- Two-year project, ended officially on Nov. 30, 2014.

## **3 – Technical Accomplishments / Results**

**Due to time limitations, I present highlights from the project**

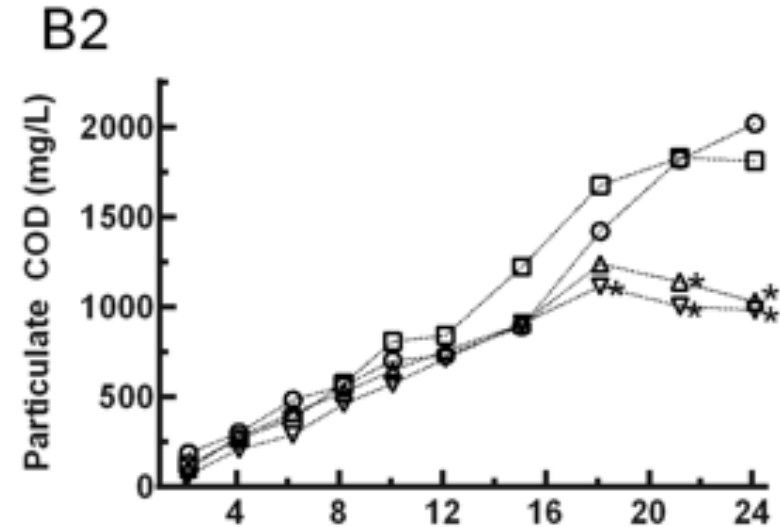
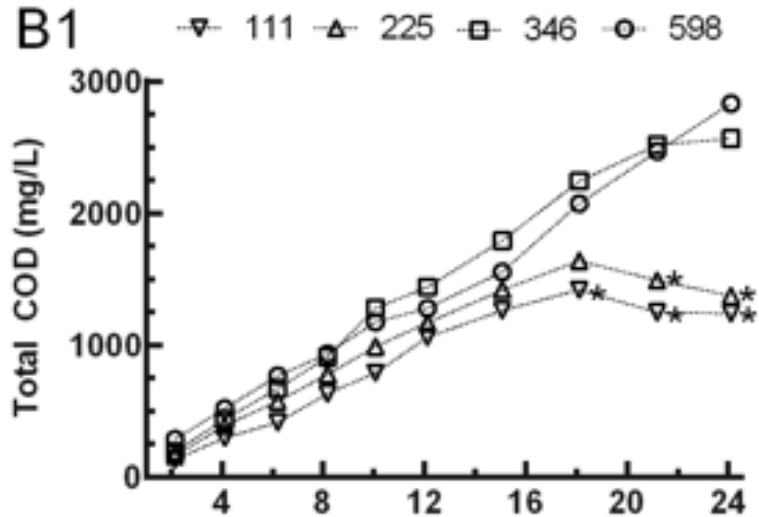
**This is ~ 20% of what we did.**

**Take-home lessons are in blue.**

# 1. *Synechocystis*



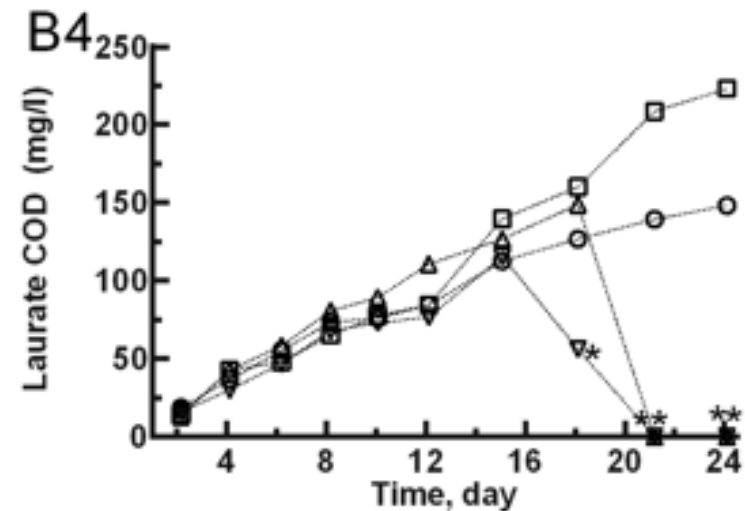
# Laurate Production by Modified *Synechocystis*



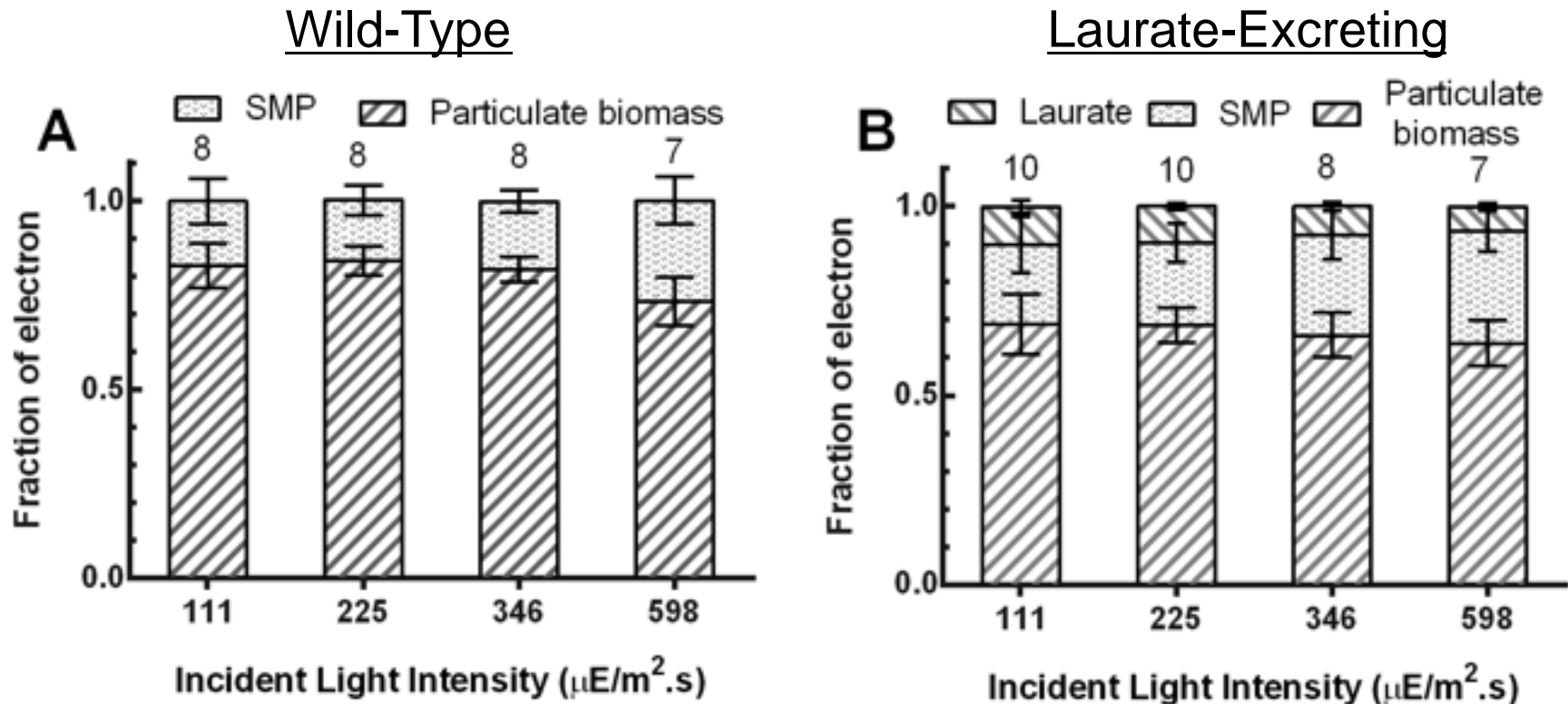
Laurate production is proportional to biomass (PCOD) growth.

Laurate is produced at less than 10% of PCOD production.

Two batch runs showed biodegradation of laurate.



# What does *Synechocystis* produce?



Both strains release at least 20% soluble microbial products (SMP), which is not a useful product.

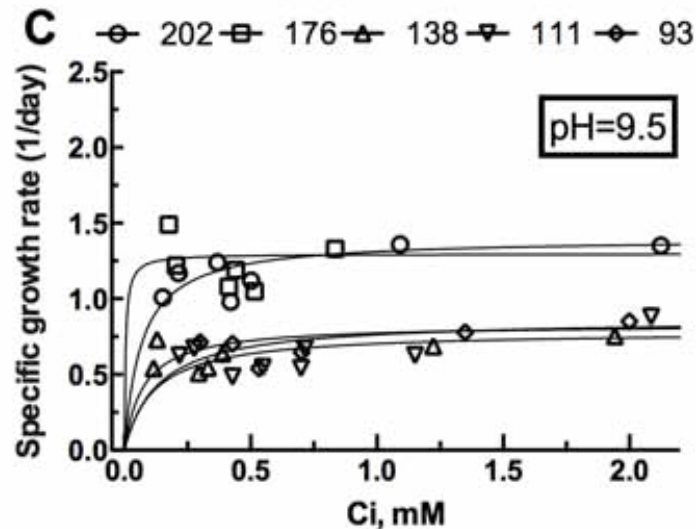
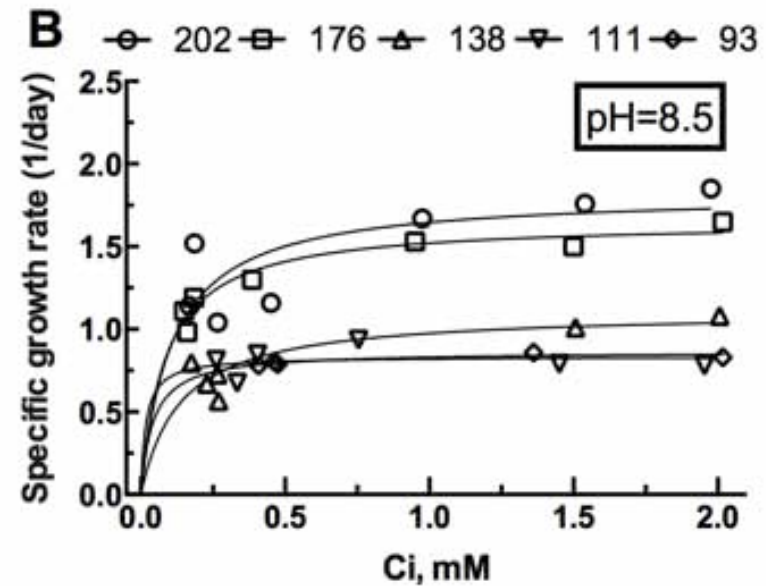
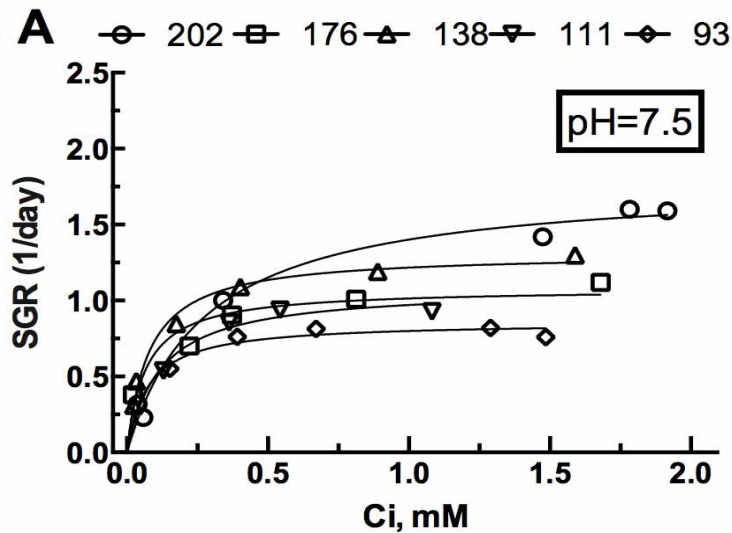
Laurate-excreting strain releases more SMP and  $\leq 10\%$  laurate.

High incident light intensity shifts partitioning to SMP and away from laurate.

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Therefore, moderate LI and get *Synechocystis* to produce less SMP!

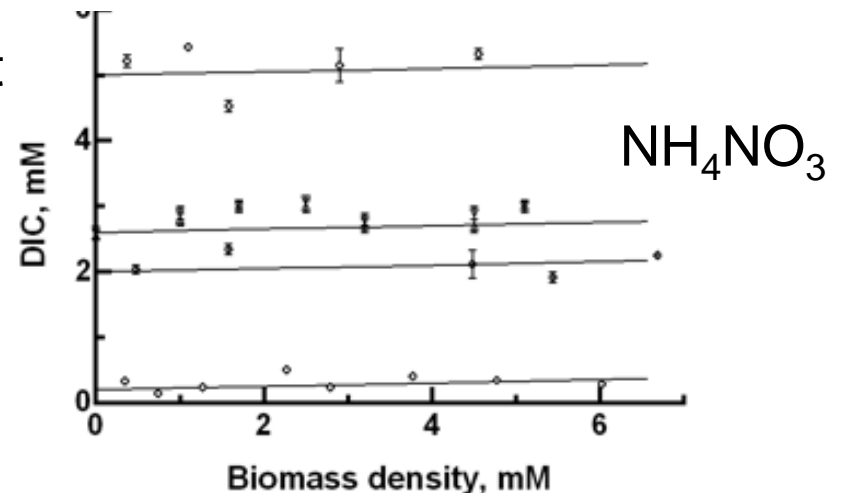
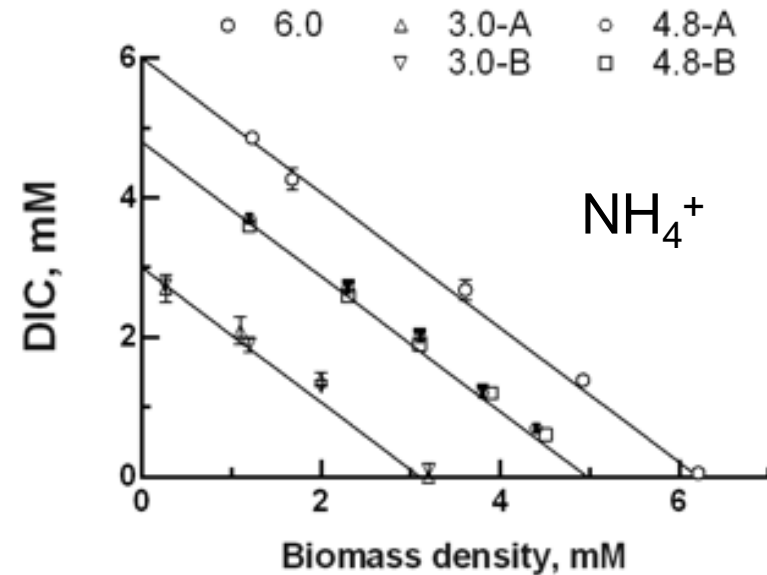
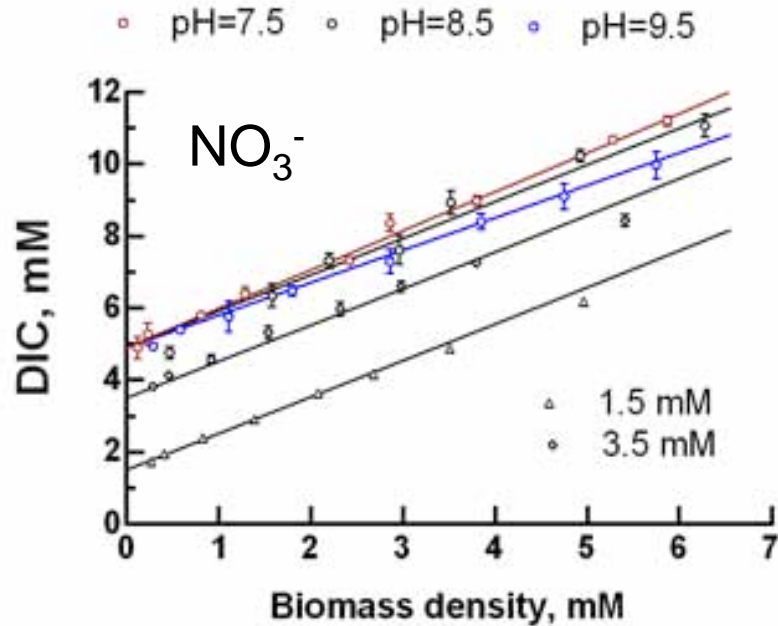
# pH and Inorganic C



For the laurate-excreting strain, the optimal pH is 8.5, and  $C_i \leq \sim 0.2$  mM (= 2.4 mgC/L) seriously slows photosynthesis.

No need to have very high  $C_i$ , but pH control is of true value.  $C_i \sim 1$  mM = 12 mgC/L and pH of 8 – 9 are good.

# Controlling pH and $C_i$

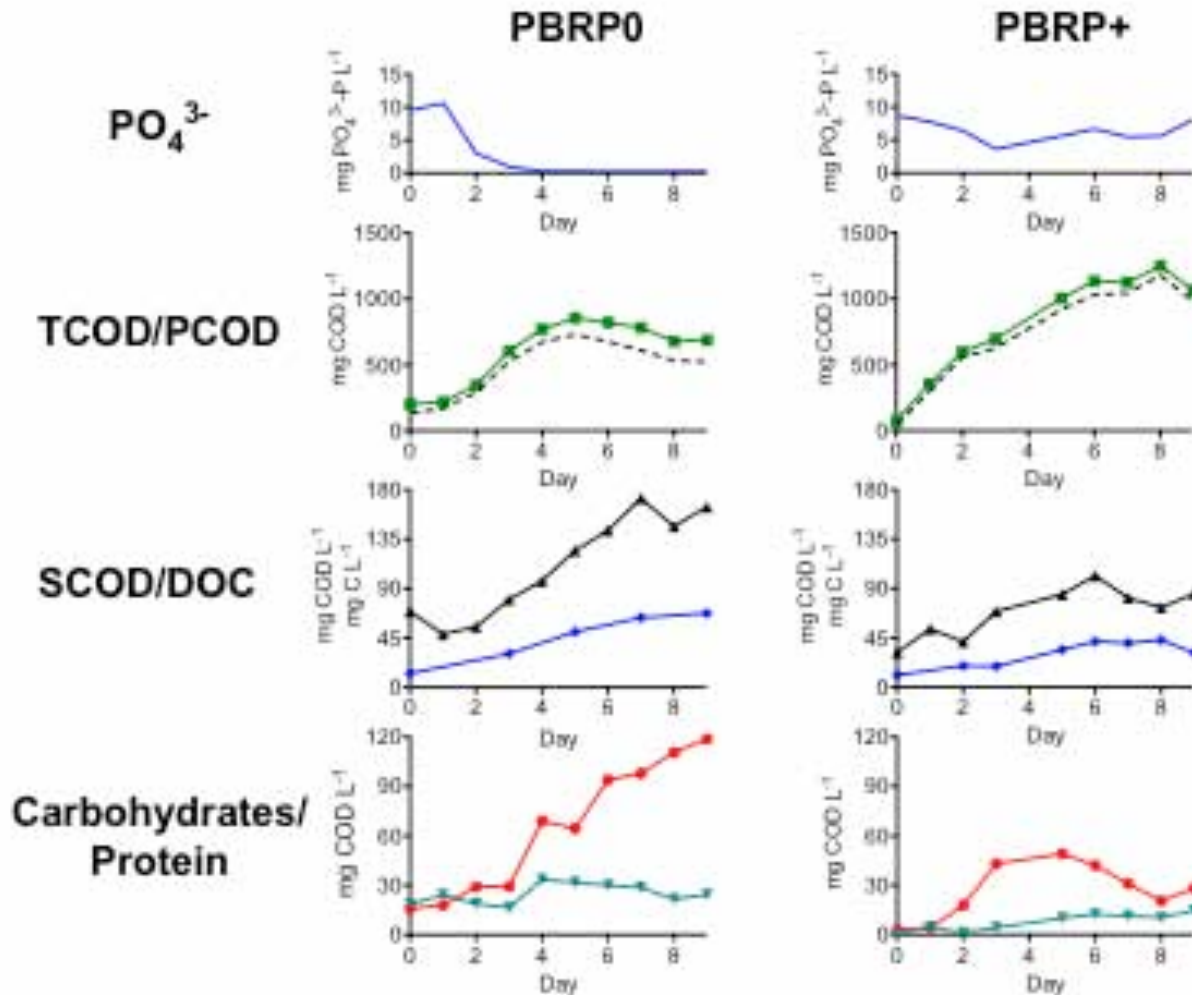


We developed and verified a model that shows that  $C_i$  is controlled by the N source in a pH-stat.

Using  $\text{NO}_3^-$  causes  $C_i$  to rise with photosynthesis.

Only  $\text{NH}_4\text{NO}_3$  holds a constant  $C_i$ .

# Phosphate Limitation

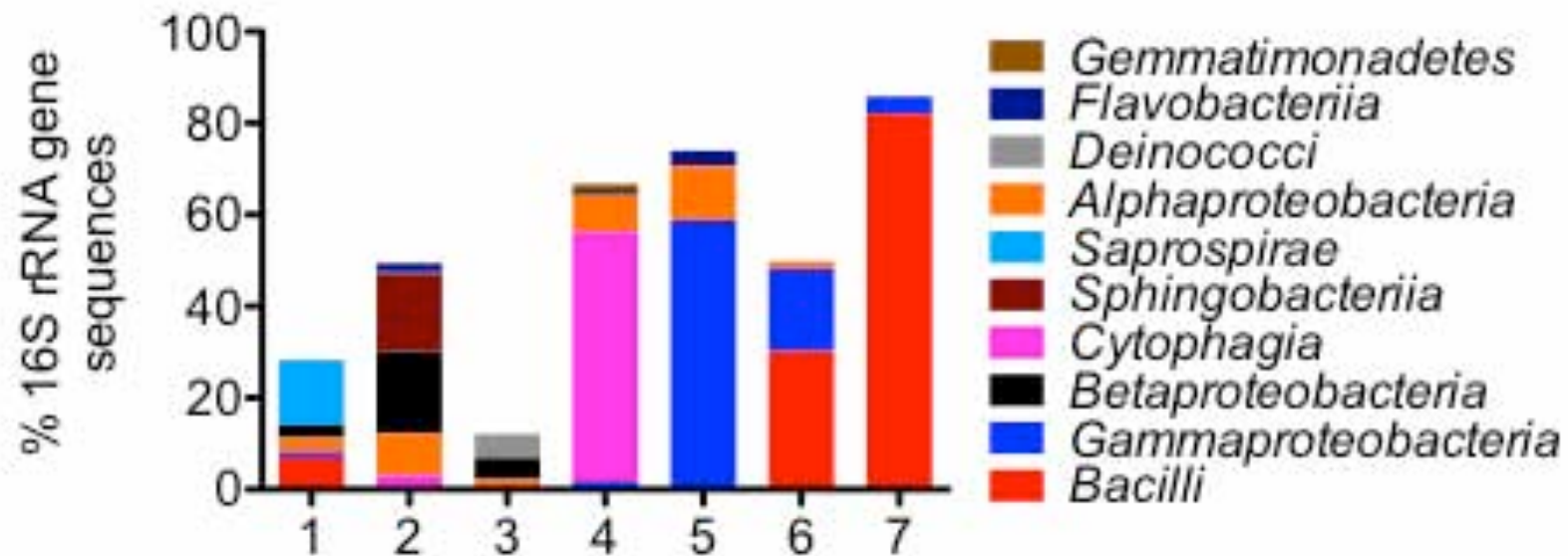


Depleting phosphate (PBR0) shows dramatic loss of biomass growth (PCOD) and increase of SMP (SCOD) and carbohydrates.

Avoid prolonged P depletion!

## 2. Heterotrophs

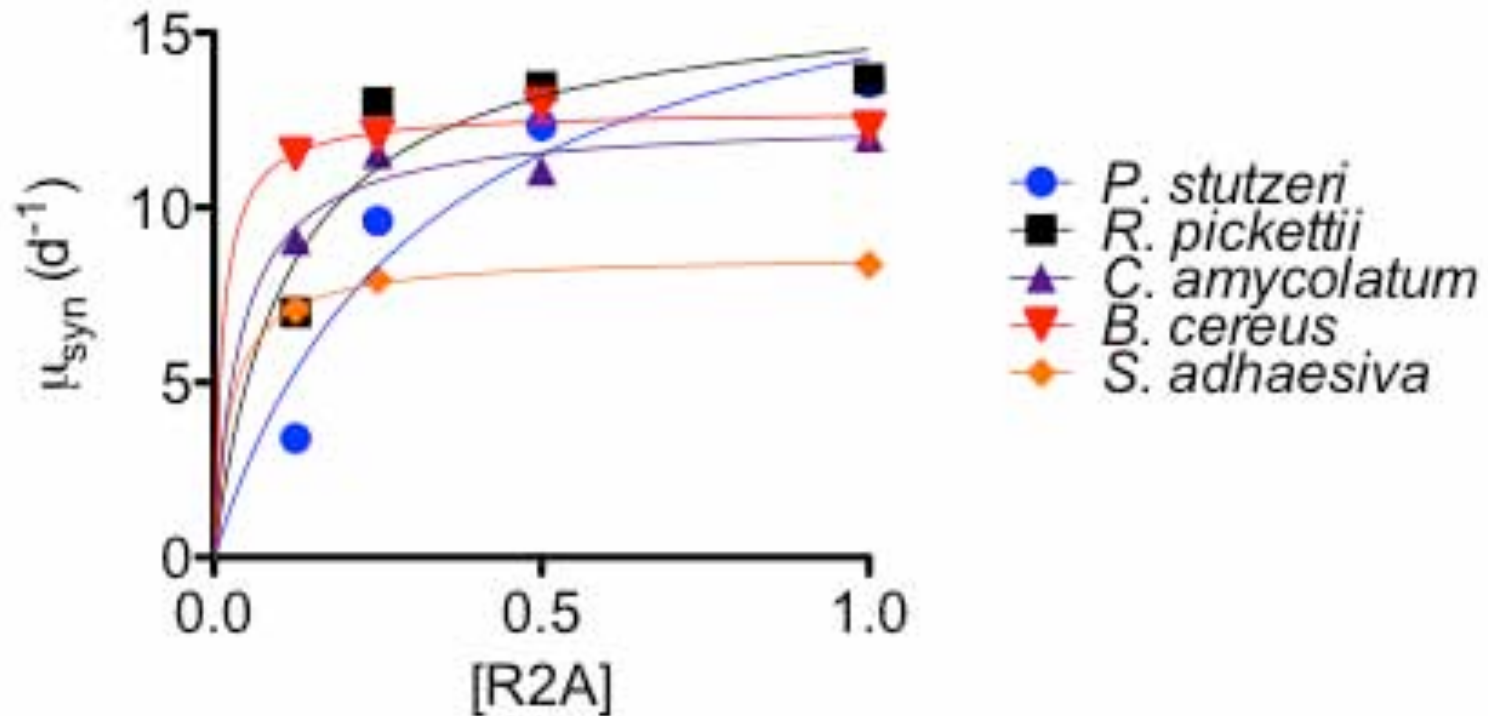
# Heterotrophs in PBRs



Phylogenetic distribution at the class level of heterotrophic bacteria in seven *Synechocystis*-based PBRs by Illumina high-throughput sequencing.

Great diversity means that the ability to live on SMP from *Synechocystis* is widespread.

# Heterotrophic Catabolism



Five PBR isolates grew rapidly on the complex medium R2A. This is a reason why they survive well in PBRs.



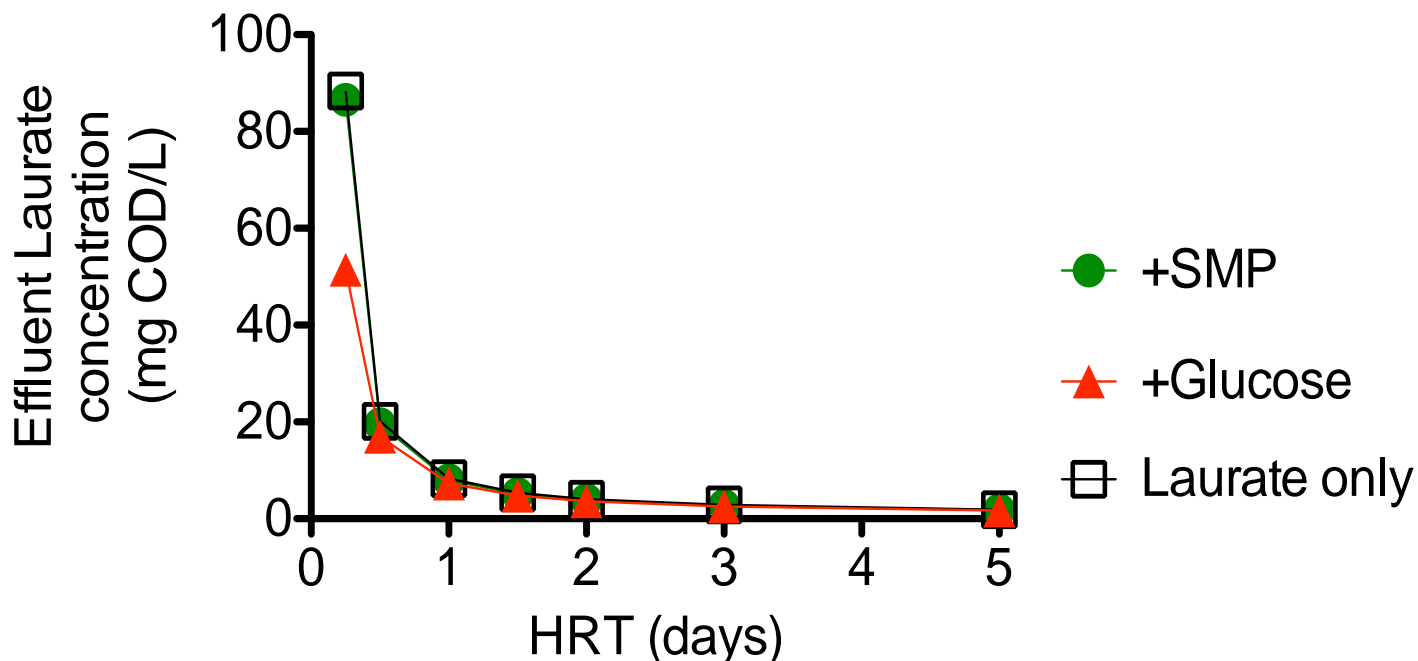
# Heterotroph Catabolism Kinetics

**Table C1.** Kinetic constants for growth of B2 and *ENR1* on acetate, laurate, and glucose.

		Acetate/Laurate		Glucose		SMP
Constant	Units	<i>P. stutzeri</i>	1-Ac	<i>P. stutzeri</i>	1-Ac	1-Ac
$Y$	mg COD/mg COD	0.51/0.5	0.37/0.52	0.52	0.51	0.3
$K$	mg COD/L	2.4/8.8	18.8/34.5	44.06	0.17	187.3
$\mu_{max}$	1/d	6.0/8.8	7.2/5.7	7.3	6.5	1.7
$q_{max}$	mg COD/mg COD*d	11.8/17.6	19.5/11.0	13.9	12.7	5.7
$S_{min}$	mg COD/L	0.04/0.1	0.7/0.62	0.62	0.003	11.7
$[\theta_x^{min}]_{lim}$	d	0.17/0.11	0.38/0.18	0.14	0.16	0.63
$K_{Pi}$	mg P/L	0.045	0.18			

Isolated strains consumed rapidly and grew well on acetate, laurate, glucose, and SMP harvested from PBRs. SMP gave slowest growth.

# Hard to Wash Out Heterotrophs



Modeled effluent laurate concentrations for a PBR with an initial laurate concentration of 100 mg COD/L and with either 10 mg COD/L SMP or 10 mg COD/L glucose.

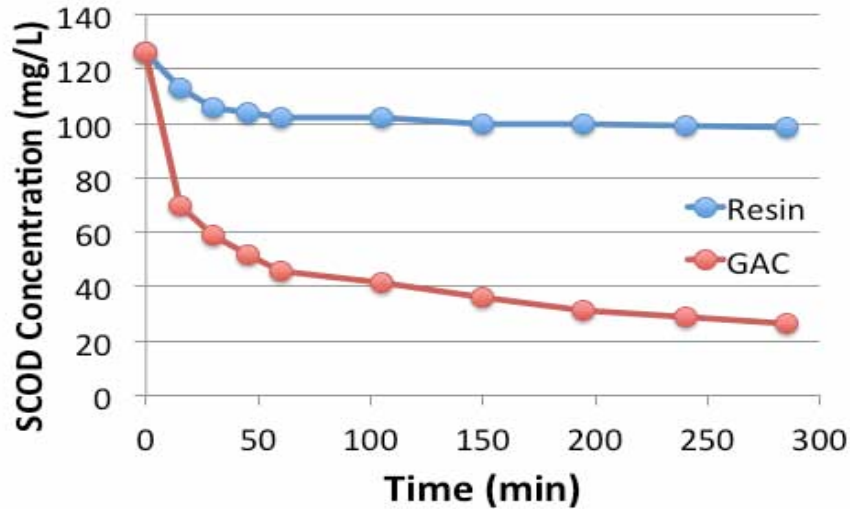
Only a very short HRT, which means a very fast specific growth rate, may be able to wash out heterotrophs and preserve laurate.

We need a PBR system able to give fast growth of *Synechocystis*.

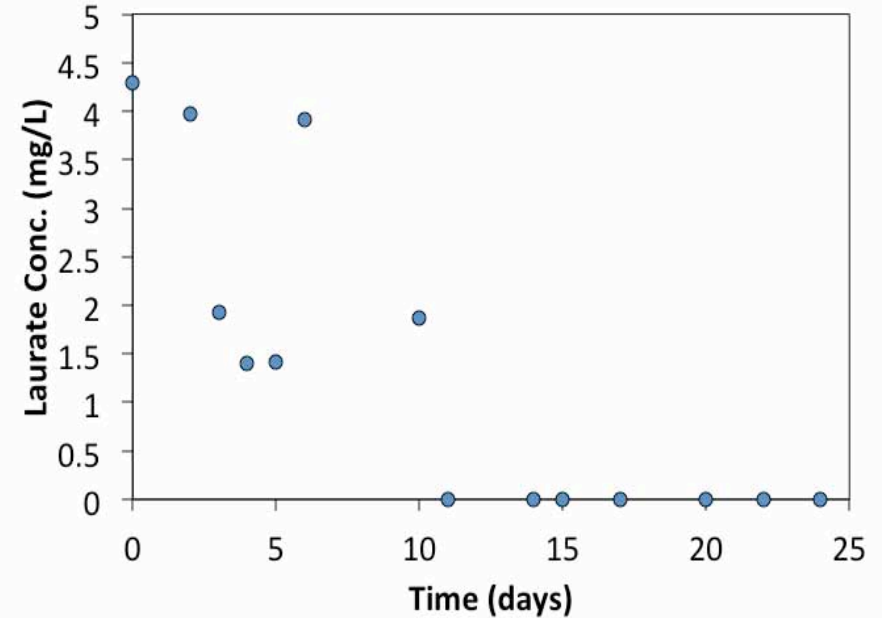
This goes well with the fact that laurate production is proportional to growth!

# 3. PBR System

# SMP Removal and Laurate Harvest



Kinetics of SMP adsorption from membrane permeate of wild-type *Synechocystis* on MP-64 resin and F-300 GAC.



Laurate concentration in the effluent for the PBR with the resin and GAC columns throughout sequencing-batch operation.

A tandem of MP-64 ion-exchange resin (for laurate harvest) and F-300 GAC (for SMP adsorption) could forestall heterotrophic biodegradation.

Need to link this with a PBR system that allows a very fast specific growth rate and laurate-production rate.

## 4 and 5 – Summary and Relevance

- Laurate-excreting *Synechocystis* generates laurate in proportion to its growth rate. **We want a fast specific growth rate.**
- *Synechocystis* partitions more fixed C to SMP than to laurate, especially for high light intensity. **We want to modify *Synechocystis* to release less SMP and to use a moderate light intensity.**
- *Synechocystis* does not need a high  $C_i$  (<12 mgC/L), but it needs a pH around 8 – 9 and no P depletion. **Control pH and use  $NH_4NO_3$ .**
- *Synechocystis*-dominated PBRs have a wide diversity of heterotrophic bacteria.
- Many heterotrophic bacteria are able to consume complex organic materials, such as in SMP.
- Harvest of laurate onto ion-exchange resin and adsorption of SMP by GAC can forestall the onset of heterotrophic biodegradation of laurate.
- **Long-term success will require a PBR able to sustain high specific growth rates (to wash out heterotrophs) and possibly modifications to *Synechocystis* so that it produces more laurate and less SMP.**

# Additional Slides

# Publications, Patents, Presentations, Awards, and Commercialization

## Publications to date (more expected)

- Nguyen, B., Rittmann, B.E. Electron partitioning into soluble organic products by wild-type and modified *Synechocystis* sp. PCC 6803. Submitted to *Algal Research*.
- Nguyen, B., Rittmann, B.E. Controlling dissolved inorganic carbon in photoautotrophic microalgae culture via the nitrogen source. Submitted to *Algal Research*
- Nguyen, B., Rittmann, B.E. Effects of inorganic carbon and pH on growth kinetics of *Synechocystis* sp. PCC 6803. Submitted to *Algal Research*
- Zevin, A.S., Masters, D.P., Rittmann, B.E., Krajmalnik-Brown, R. Phylogenetic characterization of heterotrophic bacteria isolated and enriched from PBR cultures of *Synechocystis* sp. PCC6803. Submitted to *Appl. Environ. Microb.*
- Zevin, A.S., Rittmann, B.E., Krajmalnik-Brown, R. The inoculum source has a strong influence on microbial community structure in *Synechocystis*-based PBRs. Submitted to *Algal Res.*
- Zevin, A.S., Nam, T.G., Rittmann, B.E., Krajmalnik-Brown, R. Effects of phosphate limitation on soluble microbial products and microbial community structure in semi-continuous *Synechocystis*-based photobioreactors. Submitted to *Biotechnology and Bioengineering*